

# Benzodiazepines and Metabolites from Blood and Urine by LC/MS (MRM)

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# Benzodiazepines and Metabolites from Blood and Urine by LC/MS (MRM)

## 1 INTRODUCTION

Benzodiazepines are one of the most commonly prescribed classes of drugs in the United States. They are also frequently abused. Therapeutic blood concentrations vary for benzodiazepines, depending on whether they are considered high dose benzodiazepines (e.g., diazepam) or low dose benzodiazepines (e.g., triazolam). Benzodiazepines are usually excreted into the urine as glucuronide metabolites and may persist in the urine for days after administration due to elimination half-lives that may exceed 24 hours.

## 2 SCOPE

Analyses	<input checked="" type="checkbox"/> Screening <input checked="" type="checkbox"/> Confirmation <input checked="" type="checkbox"/> Quantitation
Matrices	Blood, Urine
Analytes	7-aminoclonazepam, 7-aminoflunitrazepam, $\alpha$ -hydroxyalprazolam, $\alpha$ -hydroxymidazolam, $\alpha$ -hydroxytriazolam, alprazolam, chlordiazepoxide (qualitative only), clonazepam, desalkylflurazepam, diazepam, flunitrazepam, flurazepam (qualitative only), lorazepam, midazolam, n-desmethylflunitrazepam, nordiazepam, oxazepam, temazepam, and triazolam
Personnel	This document applies to authorized personnel who perform the described tasks, singly or in combination.

## 3 PRINCIPLE

Biological specimens are qualitatively assayed and/or quantified for benzodiazepines and their metabolites. Specimens are mixed with deuterated internal standards. Proteins are precipitated from blood before extraction. Both blood and urine samples are extracted using solid phase extraction (SPE). Analysis of extracts is by liquid chromatography/tandem mass spectrometry in the multiple reaction monitoring mode (LC/MS(MRM)).

## 4 SPECIMEN CRITERIA

This procedure uses 0.2 mL of blood (in duplicate for quantitation) or 0.4 mL of urine.

## 5 EQUIPMENT

### 5.1 Equipment

A. Centrifuge
B. Evaporator/Concentrator with Nitrogen
C. Graduated cylinders
D. Incubator
E. pH Meter
F. Pipettes
G. Vacuum or Positive Pressure Manifold
H. Volumetric flasks
I. Vortex/Mixer

#### 5.1.1 Column

Phenomenex Kinetex XB-C18 (or equivalent) analytical column (150 mm x 2.1mm x 2.6  $\mu$ )

### 5.2 Consumables

A. Centrifuge tube filters (0.45 microns, Nylon)
B. Oasis HLB 6 cc (500 mg) LP SPE cartridges
C. 12 x 75 mm test tube
D. 16 x 100 mm test tube

### 5.3 Instruments

- A. Sciex 6500+ QTRAP Mass Spectrometer
- B. Shimadzu HPLC

### 5.4 Software

Component	Software	Version
Operating System	Microsoft Windows	10 Enterprise 2016 LTSC
Mass Spectrometer	Analyst	1.7.1
Data Analysis	MultiQuant	3.0.3

## 5.5 Chemicals/Reagents

### 5.5.1 Purchased

A. Acetic acid, glacial	17 M, ≥ ACS grade
B. Ammonium acetate	99.999% purity
C. Ammonium formate	≥ Reagent Grade
D. Ammonium hydroxide, concentrated	15 M, ≥ ACS grade
E. Deionized water	18 MΩ
F. Disodium hydrogen phosphate (anhydrous)	≥ Reagent Grade
G. Formic acid	≥ Reagent Grade
H. Isopropanol (HPLC grade)	≥ HPLC grade
I. Methanol	≥ Optima Grade
J. Methylene chloride	≥ Optima Grade
K. Potassium dihydrogen phosphate	≥ Reagent Grade
L. Zinc sulfate heptahydrate	≥ Reagent Grade
M. β-Glucuronidase	>100,000 u/mL β-glucuronidase activity; from Red Abalone, <i>H. Rufescena</i> ; available from Kura Biotec

### 5.5.2 Prepared

A. Zinc Sulfate (0.2 M)	Combine 50 mL deionized water and 5.75 g of zinc sulfate heptahydrate in a 100 mL volumetric flask. Mix until dissolved. Bring to the mark with deionized water. Store in glass at room temperature. Stable for at least 6 months.
B. Zinc Sulfate in Methanol	Combine 80 mL methanol and 20 mL zinc sulfate (0.2 M) in a volumetric flask and mix well. Store in glass at room temperature. Stable for at least 2 months.
C. Potassium Phosphate Buffer (KH <sub>2</sub> PO <sub>4</sub> )	Add 4.54 g potassium phosphate monobasic to a 0.5 L volumetric flask and bring to the mark with deionized water. Store refrigerated in glass or plastic. Stable for at least three months.
D. Sodium Phosphate Buffer (Na <sub>2</sub> HPO <sub>4</sub> )	Add 11.6 g sodium phosphate dibasic anhydrous to a 1 L volumetric flask and bring to the mark with deionized water. Store refrigerated in glass or plastic. Stable for at least three months.
E. Sorensen Buffer (pH 7.4)	

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Combine 500 mL of (D) Sodium Phosphate Buffer with 200 mL of (C) Potassium Phosphate Buffer. Add (C) to decrease pH or (D) to increase pH until pH reads 7.4 with a pH meter or tight range pH strip. Store refrigerated in glass or plastic. Stable for at least three months.

**F. Ammonium Acetate Buffer (0.5 M; pH 5)**

Add 3.854 g ammonium acetate to a 100-mL volumetric flask containing about 75 mL deionized water. Mix well to dissolve. Add glacial acetic acid until pH registers between 4.5 and 5.5. Bring to volume with deionized water and mix well. Store refrigerated in glass or plastic. Stable at least three months.

**G. Methanol:Water:Ammonia (40:60:0.5)**

Combine 40 mL methanol, 60 mL deionized water and 0.5 mL ammonium hydroxide and mix well. Store at room temperature in glass. Prepare fresh daily.

**H. Methylene Chloride:Isopropanol (75:25)**

Combine 188 mL methylene chloride and 62 mL isopropanol in a 250 mL graduated cylinder and mix well. Store at room temperature in glass. Stable for at least two months.

**I. Water:Acetonitrile (90:10)**

Combine 90 mL deionized water and 10 mL acetonitrile (Optima grade) and mix well. Store at room temperature in glass. Stable for at least three months.

**J. Mobile Phase 1 (5 mM Ammonium Formate with formic acid; pH~3.5)**

Add 0.3153 g ammonium formate to a 1 L volumetric flask. Add approximately 800 mL deionized water and mix well. Add 1 mL formic acid, and fill to the line with deionized water. Store in glass at room temperature. Stable for at least one week.

**K. Mobile Phase 2 (Acetonitrile with 0.1% Formic Acid)**

Combine 1 mL formic acid and 1000 mL acetonitrile and mix well. Store in glass at room temperature. Stable for at least two months.

**L. Methanol:Water (1:1)**

Combine equal amounts methanol and water. Store in glass at room temperature. Stable for at least 12 months.

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## 5.6 Standards/Controls

### 5.6.1 Purchased

#### 5.6.1.1 Standard/Control Stock Solutions

Standard and control stock Solutions (1.0 mg/mL) of the following may be purchased from Cerilliant (Round Rock, TX), Lipomed or an equivalent supplier. Solutions may be in methanol or acetonitrile and will be stored according to the manufacturer's recommendations. Stability is determined by the manufacturer.

7-aminoclonazepam	clonazepam	n-desmethylflunitrazepam
7-aminoflunitrazepam	desalkylflurazepam	nordiazepam
$\alpha$ -hydroxyalprazolam	diazepam	oxazepam
$\alpha$ -hydroxymidazolam	flunitrazepam	temazepam
$\alpha$ -hydroxytriazolam	flurazepam	triazolam
alprazolam	lorazepam	
chlordiazepoxide	midazolam	

#### 5.6.1.2 Hydrolysis Control Stock Solution

Oxazepam-glucuronide (0.1 mg/mL) may be purchased from Cerilliant (Round Rock, TX), or an equivalent supplier; the S-isomer of the oxazepam-glucuronide is known to be cleaved by  $\beta$ -glucuronidase approximately 400 times faster than the R-isomer, making the R-isomer unsuitable as a control in this procedure.

#### 5.6.1.3 Internal Standard Stock Solutions

Internal Standard Stock Solutions (0.1 mg/mL) of the following may be purchased from Cerilliant (Round Rock, TX) or an equivalent supplier. Solutions may be in methanol or acetonitrile and will be stored according to the manufacturer's recommendations. Stability is determined by the manufacturer.

7-aminoclonazepam-d <sub>4</sub>	chlordiazepoxide-d <sub>5</sub>	midazolam-d <sub>4</sub>
7-aminoflunitrazepam-d <sub>7</sub>	clonazepam-d <sub>4</sub>	n-desmethylflunitrazepam-d <sub>4</sub>
* $\alpha$ -hydroxyalprazolam-d <sub>5</sub>	desalkylflurazepam-d <sub>4</sub>	nordiazepam-d <sub>5</sub>
$\alpha$ -hydroxymidazolam-d <sub>4</sub>	diazepam-d <sub>5</sub>	oxazepam-d <sub>5</sub>
$\alpha$ -hydroxytriazolam-d <sub>4</sub>	flunitrazepam-d <sub>7</sub>	temazepam-d <sub>5</sub>
alprazolam-d <sub>5</sub>	lorazepam-d <sub>4</sub>	triazolam-d <sub>4</sub>
*used for flurazepam		

## 5.6.2 Prepared

### 5.6.2.1 Internal Standard Prepared Solutions

#### A. Internal Standard Intermediate Solution (5 µg/mL):

Add 0.25 mL of each Internal Standard Stock Solution to a 5-mL volumetric flask and bring to the mark with methanol. Store in the freezer. Stable for at least 2 years.

#### B. Internal Standard Working Solution (500 ng/mL):

Combine 0.1 mL of the Internal Standard Intermediate Solution (5 µg/mL) and 0.9 mL methanol. Prepare fresh daily.

### 5.6.2.2 Calibration Solutions

<b>A. High Calibration Solution</b>			
Starting Solution	1	mg/mL	stock solution(s)
Starting Solution Aliquot	0.025	mL	
Diluent Volume	25	mL	methanol, volumetric flask
Resulting Concentration	1	µg/mL	storage: freezer; stability: ≥ 1 year
<b>B. Low Calibration Solution</b>			
Starting Solution	1	µg/mL	High Calibrator Solution
Starting Solution Aliquot	0.5	mL	
Diluent Volume	10	mL	methanol, volumetric flask
Resulting Concentration	50	ng/mL	storage: freezer; stability: ≥ 1 year
<b>C. Calibration Scheme</b>			
Calibrator Level	Low Cal Spike (µL)	High Cal Spike (µL)	Resulting Concentration, ng/mL (in 0.2mL of blood)
1	10		2.5
2	20		5
3	100		25
4	200		50
5		20	100
6		30	150
7		50	250
8		75	375



### 5.6.2.3 Blood Controls

Negative Control Blood is purchased from Cliniqua or another approved vendor. Storage and stability determined by manufacturer. A Negative Control Blood sample will be extracted and analyzed with every blood assay.

At least one Positive Control Blood Sample will be analyzed with each blood assay. For quantitative analyses, both levels of Positive Control Blood Samples will be analyzed in duplicate.

<b>A. High Control Solution</b>			
Starting Solution	1	mg/mL	stock solution(s)
Starting Solution Aliquot	0.025	mL	
Diluent Volume	25	mL	methanol, volumetric flask
Resulting Concentration	1	µg/mL	storage: freezer; stability: ≥ 1 year

<b>B. Low Control Solution</b>			
Starting Solution	1	µg/mL	High Control Solution
Starting Solution Aliquot	0.5	mL	
Diluent Volume	10	mL	methanol, volumetric flask
Resulting Concentration	50	ng/mL	storage: freezer; stability: ≥ 1 year

<b>C. Blood Control Scheme</b>			
	Low Control	High Control	Resulting Concentration, ng/mL
Control Level	Spike (µL)	Spike (µL)	(in 0.2mL of blood)
Negative	0	0	0
Low	20	0	5
High	0	50	250

#### 5.6.2.4 Urine Hydrolysis Solution

##### A. Hydrolysis High Control Solution (1 µg/mL Oxazepam equivalent)

To a 10-mL volumetric flask, add 0.160 mL of the Oxazepam Glucuronide Stock Standard. Bring to volume with methanol. Store frozen in glass. Stable for ≥ 1 year.

#### 5.6.2.5 Urine Controls

##### A. Urine Negative Control

Prepared in-house or purchased from an appropriate vendor. Stable for 6 months when refrigerated. A Negative Control Urine sample will be extracted and analyzed with every urine assay.

##### B. Working Standard Control Solution

Starting Solution	1	µg/mL	High Control Solution
Starting Solution Aliquot	0.5	mL	
Diluent Volume	10	mL	1:1 methanol:water, volumetric flask
Resulting Concentration	50	ng/mL	storage: freezer; stability: ≥ 1 year

##### C. Hydrolysis Working Standard Control Solution

Starting Solution	1	µg/mL	Urine Hydrolysis Solution
Starting Solution Aliquot	0.5	mL	
Diluent Volume	10	mL	1:1 methanol:water, volumetric flask
Resulting Concentration	50	ng/mL	storage: freezer; stability: ≥ 1 year

##### D. Urine Control Scheme

	WS Control Solution	High Control	Urine (mL)	Resulting Concentration, ng/mL
Control Level	Spike (µL)	Spike (µL)		
Negative	0	0	0.4	0
Low	20	0	1	1
High	0	10	1	10

Mix Low and High Controls well before removing 0.4 mL for analysis.

##### E. Hydrolysis Urine Control Scheme

	Hydrolysis WS Control Solution	Hydrolysis High	Urine (mL)	Resulting Concentration, ng/mL
--	--------------------------------	-----------------	------------	--------------------------------

		Control Solution		
Control Level	Spike (μL)	Spike (μL)		
Low Hydrolysis	20	0	1	1
High Hydrolysis	0	10	1	10

Mix Low and High Hydrolysis Controls well before removing 0.4 mL for analysis.

#### 5.6.2.6 LC/MS Performance Standard (5 ng/mL)

Add 5 μL of the Benzodiazepine High Calibrator Solution (1 μg/mL) and 10 μL of the Internal Standard Working Solution to 1 mL of Water:Acetonitrile (90:10). Store in refrigerated autosampler tray for up to one week or prepare fresh daily.

## 6 PROCEDURE

Step	Note	Reference/Lot
<b>A. Sample Preparation for Blood Specimens</b>		
1. Prepare blood calibrators as directed in Section 5.6.2.2		
High Calibration Solution	[    ]	
Low Calibration Solution	[    ]	
Negative Control Matrix	[    ]	
Pipettors Used	[    ]	
2. Prepare blood controls as directed in Section 5.6.2.3		
High Control Solution	[    ]	
Low Control Solution	[    ]	
Negative Control Matrix (at least one negative control is required)	[    ]	
Pipettors Used	[    ]	
3. Prepare case blood samples		
Add 0.2 mL of case blood sample to 12 x 75 mm test tube. (prepare in duplicate for quantitation and diluted as required for on-scale analysis)		
<b>B. Internal Standard(s) for Blood Specimens</b>		
1. Add 0.020 mL of the Internal Standard Working Solution (500 ng/mL) to each sample	[    ]	
2. Vortex well		
<b>C. Protein Precipitation and Buffering for Blood Specimens</b>		
1. Add 2 mL of zinc sulfate in methanol to each blood sample	[    ]	
2. Allow to sit for 1 minute		
3. Vortex		
4. Centrifuge samples for 5 minutes at 3000 rpm		
5. Transfer supernatant to a new, properly labeled 16 x 100 mm test tube		
6. Concentrate samples to ~ 0.4 mL under nitrogen at 60°C		
7. Add 5.5 mL of Sorenson buffer to each tube	[    ]	
8. Vortex		
9. Centrifuge samples for 1 minute at 3000 rpm		

<b>D. Sample Preparation for Urine Specimens</b>		
1. Prepare urine controls as directed in Section 5.6.2.5		
<b>Low Positive Control Urine (1 ng/mL)</b>		
Benzodiazepine Working Standard Control Solution (50 ng/mL)	[     ]	
<b>High Positive Control Urine (10 ng/mL)</b>		
High Control Solution (1 µg/mL)	[     ]	
<b>Hydrolysis Low Positive Control Urine (1 ng/mL)</b>		
Urine Hydrolysis Working Standard Control Solution (50 ng/mL)	[     ]	
<b>Hydrolysis High Positive Control Urine (10 ng/mL)</b>		
Hydrolysis High Control Solution (1 µg/mL)	[     ]	
<b>Negative Control Matrix</b>		
[     ]		
2. Prepare case urine samples		
i. Add 0.4 mL of each case urine sample into a properly labeled 16 x 100 mm test tube		
<b>E. Internal Standard(s) for Urine Specimens</b>		
1. Add 0.010 mL of the Internal Standard Working Solution (500 ng/mL) to each sample	[     ]	
2. Vortex		
<b>F. Hydrolysis for Urine Specimens</b>		
1. Add 0.6 mL ammonium acetate buffer (0.5 M, pH 5)	[     ]	
2. Add 0.1 mL β-glucuronidase	[     ]	
3. Cap and vortex		
4. Incubate for 30 minutes at 68°C		
5. Cool to room temperature		
<b>G. Protein Precipitation and Buffering for Urine Specimens</b>		
1. Add 2mL zinc sulfate in methanol, allow to sit for 1 minute	[     ]	
2. Vortex. Centrifuge samples for 5 minutes at 3000 rpm		
3. Transfer supernatant to a new 16 x 100 mm test tube		
4. Concentrate samples to ~ 1.0 mL under nitrogen at 60°C		
5. Add 5 mL of Sorenson buffer to each tube	[     ]	
6. Vortex and centrifuge samples for 1 minute at 3000 rpm		

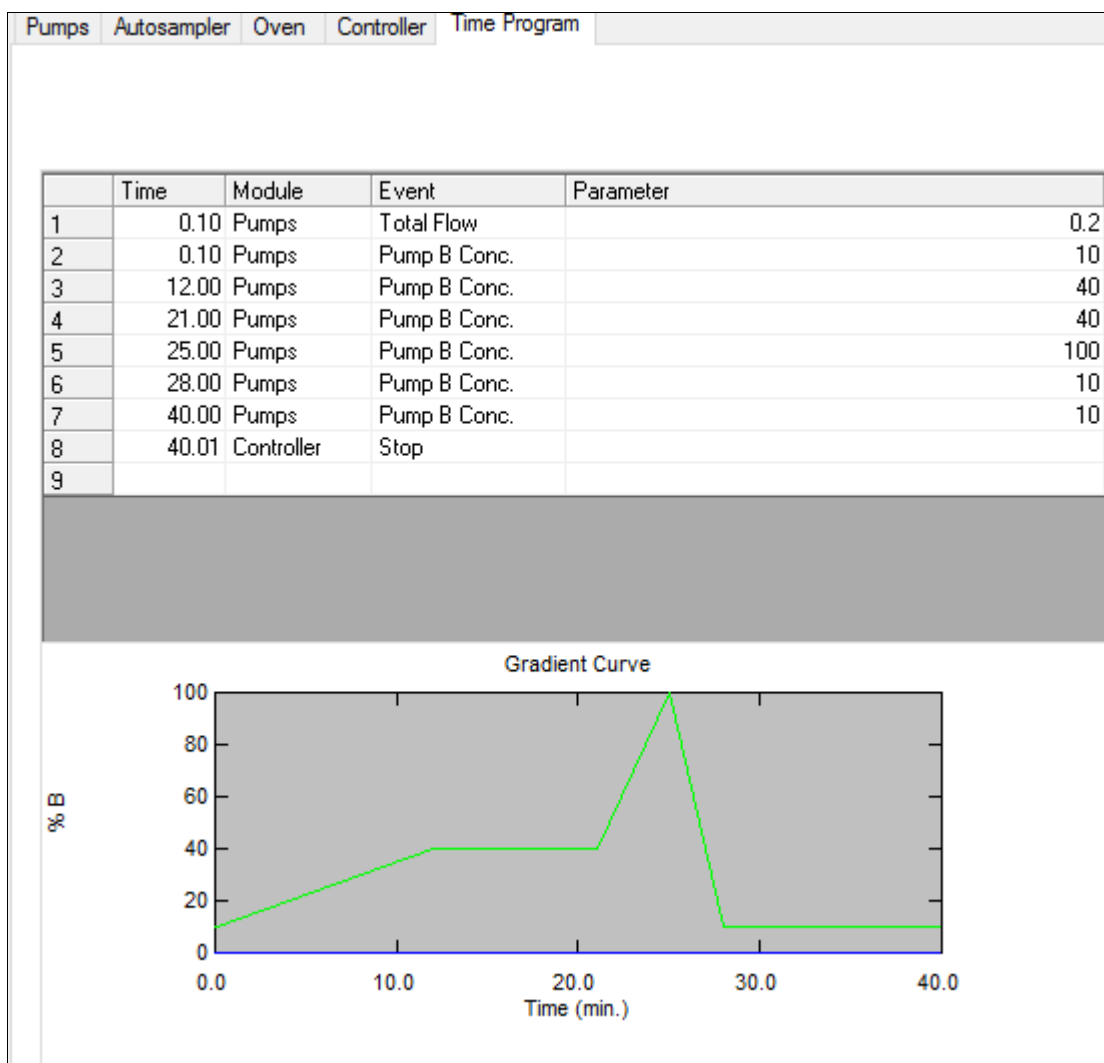
<b>H. Solid Phase Extraction (Blood and Urine Samples)</b>		
1. Condition cartridge (Oasis HLB 6 cc (500 mg) LP)	[!!!!]	
a. Add 2 mL methanol	[!!!!]	
b. Add 3 mL deionized water	[!!!!]	
2. Load samples on SPE cartridges		
3. Wash cartridge with 2 mL of methanol:water:ammonia (40:60:0.5)	[!!!!]	
4. Dry cartridges at full vacuum for 15 minutes (use vacuum manifold; positive pressure manifold does not dry sufficiently)		
5. Elute with 5 mL dichloromethane:isopropanol (75:25) under gravity	[!!!!]	
6. Evaporate eluent to dryness under nitrogen at 60°C		
<b>I. Reconstitution and Filtration</b>		
1. Blood Samples		
i. Add 0.25 mL water:acetonitrile (90:10)	[!!!!]	
2. Urine Samples		
i. Add 0.1 mL water:acetonitrile (90:10)	[!!!!]	
3. Filter extracts through 0.45 micron filters (nylon)	[!!!!]	
<b>J. Instrumental Analysis</b>		
1. LC/MS:		
i. Analyze LC/MS Performance Standard prior to batch analysis - 5 µL	[!!!!]	
ii. Blood Extracts - 5 µL		
iii. Urine Extracts - 20 µL		
2. Mobile Phase 1 (Aqueous)	[!!!!]	
3. Mobile Phase 2 (Organic)	[!!!!]	
4. LC Column	[!!!!]	

## 7 ANALYTICAL PARAMETERS

### 7.1 Liquid Chromatography

Pumps	Autosampler	Oven	Controller	Time Program
Pumping Mode: Tertiary Flow				
Total Flow:	0.2000	mL/min	Configured Pumps	
Pump B Conc:	10.0	%	Pump A: LC-20AD	
Pump C Conc:	0.0	%	Pump B: LC-20AD	
Pump B Curve:	0		Pump C: LC-20AD	
Pump C Curve:	0		Pump D: LC-20AD	
Pump D Flow:	0.0000	mL/min	Pressure Limits (Pump A, B, C)	
			Minimum: 0 psi	
			Maximum: 4500 psi	
			Pressure Limits (Pump D)	
			Minimum: 0 psi	
			Maximum: 4500 psi	

Pumps	Autosampler	Oven	Controller	Time Program																									
Model:	SIL-20AC/HT																												
Rack Type:	Undefined	<button>Detect Rack</button>																											
<input checked="" type="checkbox"/> Use Autosampler																													
Rinsing Volume:	200	uL																											
Needle Stroke:	52	mm																											
Rinsing Speed:	35	uL/sec																											
Sampling Speed:	15.0	uL/sec																											
Purge Time:	25.0	min																											
Rinse Dip Time:	0	sec																											
Rinse Mode:	After aspiration																												
<input checked="" type="checkbox"/> Enable Cooler Unit																													
Cooler Temperature:	14	°C																											
Control Vial Needle Stroke:	52	mm																											
<table border="1"><thead><tr><th>Pumps</th><th>Autosampler</th><th>Oven</th><th>Controller</th><th>Time Program</th></tr></thead><tbody><tr><td>Model:</td><td colspan="4">CTO-20AC</td></tr><tr><td><input checked="" type="checkbox"/> Enable Oven</td><td colspan="4"></td></tr><tr><td>Oven Temperature:</td><td>23</td><td>C</td><td colspan="2"></td></tr><tr><td>Maximum Temperature:</td><td>85</td><td>C</td><td colspan="2"></td></tr></tbody></table>					Pumps	Autosampler	Oven	Controller	Time Program	Model:	CTO-20AC				<input checked="" type="checkbox"/> Enable Oven					Oven Temperature:	23	C			Maximum Temperature:	85	C		
Pumps	Autosampler	Oven	Controller	Time Program																									
Model:	CTO-20AC																												
<input checked="" type="checkbox"/> Enable Oven																													
Oven Temperature:	23	C																											
Maximum Temperature:	85	C																											



## 7.2 Mass Spectrometry

MS **Advanced MS**

Experiment: 1

Scan type: MRM (MRM)

Polarity:  Positive  Negative

Scheduled MRM:  Enabled  Basic  Advanced [Import List](#)

Period Summary

Duration: 30.036 (min) Delay Time: 0 (sec)

Cycles: 901 Cycle: 2.0002 (sec)

Scheduled Ionization:  Scheduled Ionization

Start Time: 0 (min) Stop Time: 0 (min)



Source/Gas	Compound
Ion Source:	Turbo Spray IonDrive
Curtain Gas (CUR)	35.0
Collision Gas (CAD)	Low
IonSpray Voltage (IS)	5500.0
Temperature (TEM)	670.0
Ion Source Gas 1 (GS1)	50.0
Ion Source Gas 2 (GS2)	50.0

Apply the following parameters to all other experiments of the same polarity:

Source/Gas     Compound

Source/Gas	Compound
Declustering Potential (DP)	131.0
Entrance Potential (EP)	10.0
Collision Energy (CE)	33.0
Collision Cell Exit Potential (CXP)	14.0

Apply the following parameters to all other experiments of the same polarity:

Source/Gas     Compound

MS	Advanced MS
Resolution Q1:	Unit
Resolution Q3:	Unit
Intensity threshold (total count):	0
Settling time:	0 (ms)
Pause between mass ranges:	5.007 (ms)

### 7.3 Analyte Specific Parameters

Q1 Mass (Da)	Q3 Mass (Da)	Dwell Time (msec)	Analyte	DP(v)	CE(v)	CXP(v)
286.233	222.200	12.4	7_aminoclonazepam_1	131	33	14
286.233	250.200	12.4	7_aminoclonazepam_2	131	29	14
286.233	195.200	12.4	7_aminoclonazepam_3	131	43	12
288.219	222.200	12.4	7_aminoclonazepam_4	100	33	12
290.218	226.200	12.3	7_aminoclonazepam_d4	126	35	14
284.285	227.300	14.1	7_aminoflunitrazepam_1	151	35	14
284.285	240.300	14.1	7_aminoflunitrazepam_2	151	45	14
284.285	135.000	14.1	7_aminoflunitrazepam_3	151	35	14
291.285	138.000	13.9	7_aminoflunitrazepam_d7	121	39	16
325.250	297.200	19.4	alpha_hydroxyalprazolam_1	106	35	16
325.250	216.100	19.4	alpha_hydroxyalprazolam_2	106	53	24
327.247	299.200	19.4	alpha_hydroxyalprazolam_3	181	35	16
330.273	302.300	19.2	alpha_hydroxyalprazolam_d5	136	37	16
342.211	203.000	16.6	alpha_hydroxymidazolam_1	71	35	12
342.211	168.100	16.6	alpha_hydroxymidazolam_2	71	49	18
344.216	205.000	16.6	alpha_hydroxymidazolam_3	111	35	12
346.232	203.000	16.6	alpha_hydroxymidazolam_d4	151	37	12
359.211	176.000	19.3	alpha_hydroxytriazolam_1	151	37	20
359.211	331.300	19.3	alpha_hydroxytriazolam_2	151	37	18
359.211	239.200	19.3	alpha_hydroxytriazolam_3	151	61	14
361.195	333.200	19.3	alpha_hydroxytriazolam_4	100	37	18
363.229	176.000	19.2	alpha_hydroxytriazolam_d4	176	37	12
309.266	205.100	23.2	Alprazolam_1	130	45	12
309.266	281.200	23.2	Alprazolam_2	130	35	16
311.237	283.200	23.2	Alprazolam_3	126	35	16
314.253	279.300	23.0	Alprazolam_d5	131	35	16
300.217	227.100	14.9	chlordiazepoxide_1	91	33	18
300.217	241.051	14.9	chlordiazepoxide_2	91	31	16
300.217	165.100	14.9	chlordiazepoxide_3	91	63	18
305.133	232.100	14.8	chlordiazepoxide_d4	81	33	14
316.194	270.200	22.5	clonazepam_1	146	35	16
316.194	214.100	22.5	clonazepam_2	146	49	12
318.184	272.200	22.5	clonazepam_3	141	35	14
320.238	274.200	22.3	clonazepam_d4	191	35	16
289.173	226.200	24.3	Desalkylflurazepam_1	36	39	12
289.173	179.100	24.3	Desalkylflurazepam_2	36	59	20
289.173	140.004	24.3	Desalkylflurazepam_3	36	39	16
291.132	226.200	24.3	Desalkylflurazepam_4	111	39	14
293.205	230.200	24.1	Desalkylflurazepam_d4	56	39	14

Q1 Mass (Da)	Q3 Mass (Da)	Dwell Time (msec)	Analyte	DP(v)	CE(v)	CXP(v)
285.086	193.200	26.9	diazepam_1	126	47	22
285.086	154.100	26.9	diazepam_2	126	35	18
287.214	193.100	26.9	diazepam_3	141	43	22
290.203	198.200	26.8	diazepam_d5	46	45	24
314.220	268.200	25.3	flunitrazepam_1	106	35	14
314.220	239.200	25.3	flunitrazepam_2	106	45	14
314.220	183.100	25.3	flunitrazepam_3	106	67	20
321.295	275.300	24.9	flunitrazepam_d7	101	37	16
388.577	315.200	17.1	flurazepam_1	56	31	18
388.577	288.200	17.1	flurazepam_2	56	35	16
390.313	317.200	17.1	flurazepam_3	121	31	18
321.203	275.100	21.5	Lorazepam_1	91	33	14
321.203	229.200	21.5	Lorazepam_2	91	43	12
323.186	277.200	21.5	Lorazepam_3	96	29	16
327.203	281.100	21.4	Lorazepam_Cl_d4	146	27	6
326.205	249.100	16.8	Midazolam_1	176	51	14
326.205	291.300	16.8	Midazolam_2	176	37	16
326.205	222.300	16.8	Midazolam_3	176	63	28
328.190	291.200	16.8	Midazolam_4	150	37	16
330.237	253.100	16.7	midazolam_d4	126	53	14
300.173	198.100	20.4	N_desmethylflunitrazepam_1	91	53	22
300.173	254.200	20.4	N_desmethylflunitrazepam_2	91	37	14
300.173	225.100	20.4	N_desmethylflunitrazepam_3	91	47	12
304.211	258.200	20.3	N_desmethylflunitrazepam_d4	116	33	16
271.258	208.300	23.5	Nordiazepam_1	141	39	12
271.258	165.300	23.5	Nordiazepam_2	141	37	20
271.258	140.025	23.5	Nordiazepam_3	141	37	16
276.382	213.200	23.1	Nordiazepam_d5	26	39	12
287.231	241.300	20.2	Oxazepam_1	161	31	14
287.231	231.200	20.2	Oxazepam_2	161	31	12
289.213	243.200	20.2	Oxazepam_3	106	29	14
292.196	246.200	20.1	Oxazepam_d5	101	33	14
301.266	255.100	25.3	Temazepam_1	101	29	22
301.266	177.000	25.3	Temazepam_2	101	51	20
303.251	257.200	25.3	Temazepam_3	71	29	16
306.215	260.200	25	Temazepam_d5	81	33	16
343.213	239.200	24.3	Triazolam_1	36	55	12
343.213	308.300	24.3	Triazolam_2	36	35	18
345.212	241.100	24.3	Triazolam_3	36	55	14
347.216	243.300	24.1	Triazolam_d4	166	57	14

## 8 DATA ANALYSIS

### 8.1 Decision Criteria

#### 8.1.1 LC/MS Performance Standard Decision Criteria

Peaks should show good chromatographic fidelity, with reasonable peak shape, width, and resolution. The authorized individual should ensure that the peaks entirely elute within their MRM windows, and adjust the MRM window times, if necessary.

#### 8.1.2 Unknown Sample Decision Criteria

The following criteria are used as guidelines in determining the acceptability of the data produced in this assay.

##### 8.1.2.1 *Batch Acceptance*

No analytes of interest should be detected in the Negative Control. For this purpose, analytes of interest are defined as any analytes that are being reported for this batch.

Each of the analytes in the Positive Control should be detected in the LC/MS data. High and Low Positive Controls should fall within  $\pm 20\%$  of the target value. See TOX-101 for further guidance.

Oxazepam should be detected in the Urine Hydrolysis Positive Controls. The peak area of the first transition for oxazepam should be within  $\pm 50\%$  of the area of oxazepam in the Low and High Positive Control Urine samples.

##### 8.1.2.2 *Unknown Sample Criteria*

Each of the Internal Standards should be detectable in the LC/MS data.

##### 8.1.2.2.1 *Chromatography*

The peak of interest should show good chromatographic fidelity, with reasonable peak shape, width, and resolution. In order to be determined acceptable, a chromatographic peak in an unknown sample should compare favorably to a chromatographic peak of the same analyte in a known sample analyzed on the same system in the same or subsequent analytical runs.

##### 8.1.2.2.2 *Retention Time*

The retention time of the peak should be within  $\pm 2\%$  of the retention time (relative or absolute, as appropriate) obtained from injection of a reference standard, an extracted Positive Control, or an appropriate deuterated analog.

##### 8.1.2.2.3 *Signal-to-Noise*

To justify the existence of a peak, its baseline signal to peak-to-peak noise ratio should exceed 10 when using the Analyst software. Further, the baseline signal for the peak of interest should be at least 10-fold greater than that for any observed peak at similar retention time in a Negative Control or solvent blank injected just prior to the sample.

#### 8.1.2.2.4 Mass Spectrometry

At least three independent MS/MS experiments are conducted for each analyte. Two ion ratios are calculated for each analyte. The mass spectrum of the analyte of interest should match that of a reference standard, extracted calibrator, or an extracted Positive Control. See TOX-104 for further guidance.

## 8.2 Calculations

### 8.2.1 Calibration

<b>Model</b>	Linear
<b>Weighting</b>	1/x <sup>2</sup>

Refer to TOX-101 for further guidance.

### 8.2.2 Software

Calculations may be performed by one or more of the following software packages:

- A. Sciex
  - 1. Analyst
  - 2. Multiquant
  - 3. Report Builder
- B. Microsoft
  - 1. Excel

## 9 REPORTING

### 9.1 Measurement Uncertainty

Refer to CHEM-100 and TOX-101.

## 10 CORRECTIVE MEASURES

Refer to TOX-101 for guidance on action steps in the event of a quality control failure.

## 11 PERFORMANCE CHARACTERISTICS

### 11.1 LOD

Blood: 1.25 ng/mL (or lower)

Urine: 0.5 ng/mL (or lower)

### 11.2 LOQ

Blood: 2.5 ng/mL

### 11.3 Linearity

Blood: 2.5 to 375 ng/mL

### 11.4 Bias

n=15 for values in table

	Bias (%; at 5 ng/mL)	Bias (%; at 100 ng/mL)	Bias (%; at 250 ng/mL)
7-aminoclonazepam	-3.63	2.06	3.65
7-aminoflunitrazepam	2.71	5.89	1.32
α-OH midazolam	3.93	3.76	0.44
α-OH alprazolam	3.83	3.88	-1.22
diazepam	5.21	4.54	-2.80
clonazepam	0.39	3.69	1.41
alprazolam	6.84	6.45	-1.85
flunitrazepam	1.23	2.09	-0.17
desalkylflurazepam	2.71	2.42	0.36
lorazepam	2.04	2.89	0.69
n-desmethylflunitrazepam	0.09	2.11	-0.78
midazolam	0.03	2.17	1.01
nordiazepam	2.29	4.72	-2.76
oxazepam	0.52	0.97	-1.00
temazepam	5.29	5.26	-2.21
triazolam	7.57	5.43	-3.56
α-OH triazolam	2.31	4.94	-0.14

### 11.5 Precision

As both repeatability and intermediate precision; n=15 for all values in tables

	Repeatability (%; at 5 ng/mL)	Repeatability (%; at 100 ng/mL)	Repeatability (%; at 250 ng/mL)
7-aminoclonazepam	7.20	9.71	8.96
7-aminoflunitrazepam	7.29	6.49	11.09
α-OH midazolam	5.45	3.22	4.26
α-OH alprazolam	4.95	2.78	4.22
diazepam	5.26	2.34	3.33
clonazepam	4.72	2.47	3.86
alprazolam	4.83	3.66	2.85
flunitrazepam	4.54	3.02	2.76
desalkylflurazepam	4.79	4.03	4.01
lorazepam	6.79	3.28	3.12
n-desmethylflunitrazepam	3.94	3.62	3.76
midazolam	4.31	3.17	3.75

	Repeatability (%; at 5 ng/mL)	Repeatability (%; at 100 ng/mL)	Repeatability (%; at 250 ng/mL)
nordiazepam	5.00	2.79	3.73
oxazepam	5.08	2.65	3.08
temazepam	4.32	2.70	2.96
triazolam	4.59	3.17	1.92
$\alpha$ -OH triazolam	4.67	3.37	4.00
	Intermediate Precision (%; at 5 ng/mL)	Intermediate Precision (%; at 100 ng/mL)	Intermediate Precision (%; at 250 ng/mL)
7-aminoclonazepam	10.29	9.71	9.17
7-aminoflunitrazepam	7.29	6.49	11.65
$\alpha$ -OH midazolam	6.45	5.25	5.74
$\alpha$ -OH alprazolam	5.72	5.04	5.37
diazepam	5.36	3.94	5.95
clonazepam	4.91	4.05	5.37
alprazolam	5.63	5.04	6.66
flunitrazepam	5.04	4.74	4.47
desalkylflurazepam	5.44	4.64	6.60
lorazepam	7.67	7.38	7.53
flurazepam	10.45	7.50	6.29
n-desmethylflunitrazepam	5.40	4.50	4.81
midazolam	5.29	5.09	6.33
nordiazepam	5.40	5.20	5.40
oxazepam	5.68	3.29	3.16
temazepam	5.52	5.84	5.45
triazolam	5.11	5.21	5.34
$\alpha$ -OH triazolam	4.91	5.83	6.22

### 11.6 Carryover

No significant carryover detected for the linear range established.

### 12 LIMITATIONS

Oxazepam may be unstable in methanol.

### 13 SAFETY

Take standard precautions for the handling of chemicals and biological materials. Refer to the *FBI Laboratory Safety Manual* for guidance.

## 14 REVISION HISTORY

Revision	Issued	Changes
08	06/01/2023	Complete document reformat.  2 – Clarified chlordiazepoxide suitable for qualitative analysis only  5.6.1.2, 5.6.2.5.c, 5.6.2.5.e – updated to use d0-oxazepam glucuronide versus d5 version (which is no longer available) as a hydrolysis process control  6-j – specified injection volumes for extracts  8.1.2.1 – updated guidance for evaluating hydrolysis control to account for change from d5 to d0 glucuronide